



Revista Española de Nutrición Humana y Dietética

Spanish Journal of Human Nutrition and Dietetics

ARTÍCULO DE REVISIÓN – *versión post-print*

Esta es la versión aceptada. El artículo puede recibir modificaciones de estilo y de formato.

Does microbiota influence the risk of childhood obesity?

¿Influye la microbiota en el riesgo de obesidad infantil?

Rita Cristina Sanches Oliveira^{a*}, Pedro Miguel Barata de Silva Coelho^a, María del Carmen Lozano Estevan^b.

^a Faculdade de Ciências da Saúde, Universidade Fernando Pessoa. Porto, Portugal.

^b Departamento de Farmacia, Universidad Alfonso X el Sabio. Madrid, España.

* ritao@ufp.edu.pt

Recibido: 31/03/2017; Aceptado: 10/12/2017; Publicado: 21/05/2018

CITA: Oliveira R, Coelho PMBS, Lozano Estevan MC. Does microbiota influence the risk of childhood obesity?. Rev Esp Nutr Hum Diet. 2017; 22(2). doi: 10.14306/renhyd.22.2.389 [ahead of print]

La Revista Española de Nutrición Humana y Dietética se esfuerza por mantener a un sistema de publicación continua, de modo que los artículos se publiquen antes de su formato final (antes de que el número al que pertenecen se haya cerrado y/o publicado). De este modo, intentamos poner los artículos a disposición de los lectores/usuarios lo antes posible.

The Spanish Journal of Human Nutrition and Dietetics strives to maintain a continuous publication system, so that the articles are published before its final format (before the number to which they belong is closed and/or published). In this way, we try to put the articles available to readers/users as soon as possible.

ABSTRACT

Childhood obesity is associated to incremented risk of developing diseases such as diabetes, cardiovascular diseases, or cancer, later in life. Several factors affect infant weight gain such as genetics, maternal lifestyle, and other environmental factors. Perinatal period is considered to be the most important one to when defining metabolic programming of the future adult. Several previous researches have discussed the role that gut microbiota might play on obesity risk and its development between 3-5 years old. Again, perinatal period is crucial to define quantity and diversity of a healthy intestinal microbiota. Maternal diet/BMI, delivery mode, antibiotic exposure and breastfeeding are some of the processes that will determine a favorable gut microbiota. Functions of gut microbiota, mostly by producing short-chain fatty acids as metabolites, include regulation of metabolism and immune system of the host, which may be compromised in case of dysbiosis. This review pretends to evaluate the state of the art concerning infant obesity and the role of gut microbiota. Despite the large amount of scientific publications, there is still much work to do regarding the clarification of mechanisms and the possible therapy for childhood obesity.

Keywords: Obesity; Gastrointestinal Microbiome; Pediatric Obesity; Breast Feeding; Dysbiosis.

RESUMEN

La obesidad infantil se asocia con el incremento del riesgo de desarrollar futuras enfermedades como la diabetes, las enfermedades cardiovasculares o el cáncer. Varios factores afectan la ganancia de peso infantil, como la genética, el estilo de vida materno y otros factores ambientales. El período perinatal es considerado como el más importante a la hora de definir la programación metabólica del futuro adulto. Varias investigaciones previas han discutido el rol que podría tener la microbiota intestinal en el riesgo de obesidad y su desarrollo entre los 3 y 5 años. Una vez más, el período perinatal es crucial para definir la cantidad y la diversidad de una microbiota intestinal saludable. La dieta materna, el tipo de parto, la exposición a los antibióticos y la lactancia materna son algunos de los procesos que determinarán una microbiota intestinal favorable. Las funciones de la microbiota intestinal, principalmente mediante la producción de ácidos grasos de cadena corta como metabolitos, incluyen la regulación del metabolismo y el sistema inmunológico del huésped, que pueden estar comprometidos en caso de disbiosis. Esta revisión pretende evaluar el estado del arte en relación con la obesidad infantil y el papel de la microbiota intestinal. A pesar de la gran cantidad de publicaciones científicas, todavía hace falta aclarar los mecanismos y la posible terapia para la obesidad infantil.

Palabras clave: Obesidad; Microbioma Gastrointestinal; Obesidad Pediátrica; Lactancia Materna; Disbiosis.

INTRODUCTION

World Health Organization (WHO) has stated the following facts about obesity¹:

- Obesity is defined as an abnormal or excessive fat accumulation that may impair health and results from an energy imbalance between calories consumed and calories expended;
- Overweight and obesity are linked to more deaths worldwide than underweight;
- Globally there are more people who are obese than underweight – this occurs in every region except parts of sub-Saharan Africa and Asia;
- In 2014, more than 1.9 billion adults, 18 years and older, were overweight and of these over 600 million were obese;
- In 2014, 41 million children under the age of 5 were overweight or obese.

Obesity is considered an epidemic disease escalating in all population groups in developed and developing countries. The prevalence of overweight and obesity combined has risen by 27.5% for adults and 47.1% for children between 1980 and 2013². Some explanations for the epidemiological obesity were proposed including increases in energy intake, changes in the composition of diet, reduced physical activity, and changes in the gut microbiome². Excessive weight gain in infancy is associated with persistence of high weight status and later obesity, resulting in an incremented risk to develop diseases such as diabetes, cardiovascular diseases, musculoskeletal disorders, cancer and mortality^{1,3}. Obese children can also present breath difficulties, increased risk of fractures, hypertension, insulin resistance and psychological disorders¹. Infant obesity and severe obesity has increased over the recent decades and despite this increase appears to slow down, the prevalence of child obesity is still too high worldwide⁴. Woo Baidal et al. concluded in their systematic review from prospective studies with scientific evidence, that the main factors affecting childhood obesity are higher maternal pre-pregnancy body mass index (BMI), prenatal tobacco exposure, maternal excess gestational weight gain, high infant birth weight, and accelerated infant weight gain. They also found that the critical period is from conception through 2nd years old⁵.

With different degrees of evidence, there are some important issues that appear to determine the risk of obesity in childhood: genetics⁶⁻⁸ and epigenetics^{3,9}; in uterus environment and maternal health^{4,10}; growth acceleration in the 1st six months^{10,11}; metabolic programming of endocrine response¹¹⁻¹⁴; breastfeeding and infant formulas¹⁵⁻²⁴; introduction of solid food²⁵; and intestinal microbiota¹². Intestinal microbiota is defined early in life and recent studies suggested its relation to later obesity risk and other diseases. The intestinal microbiota influences energy balance producing short-chain fatty acids (SCFA) from polysaccharides digestion. Different bacteria have

diverse modes of influencing absorption and storage of energy and several factors define colonization in infants¹².

To prevent childhood obesity, it is imperative to work on these factors, particularly those who can be affected by maternal or caregiver behavior: maternal BMI, breastfeeding human milk (breast vs bottle), formula composition, feeding practices for introduction of solids, early nutritional education with impact on metabolism routes, taste preferences and food choices in the future²⁶. The intestinal microbiota is a new insight that has emerged in recent investigations as a modulating factor of obesity. Knowing its possible role as a risk factor in childhood obesity is the main goal of this review.

MECHANISMS OF ACTION OF THE INTESTINAL MICROBIOTA

The impact of microbiota metabolism in human body has been discovered as more studies have been published linking intestinal microbiota and some developed pathologies, in germ-free mice and in humans. Alterations in microbiota populations are related to the development of inflammatory and metabolic diseases like inflammatory bowel disease, obesity, type 2 diabetes, atherosclerosis, allergy, and cancer^{27,28}.

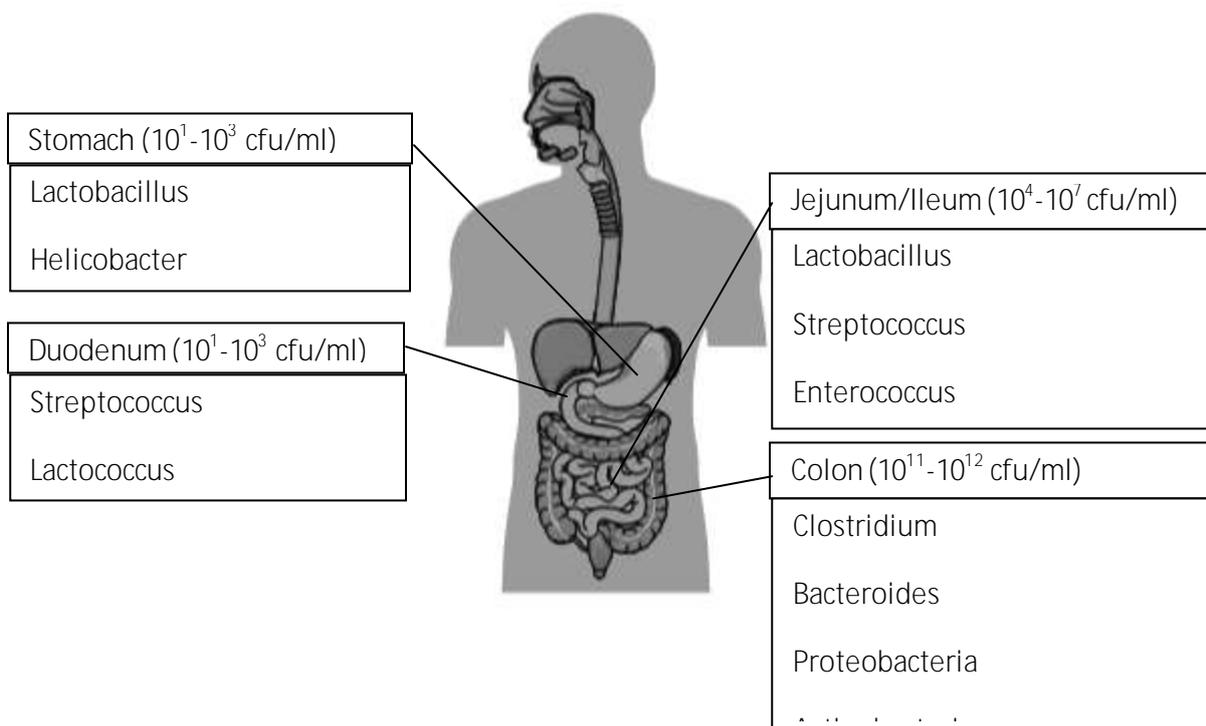
Gut microbiota ferment complex polysaccharides and residual proteins that cannot be digested by human enzymes to SCFA, branched-chain fatty acids, gases, and other metabolites. Acetate (C2), propionate (C3) and butyrate (C4) are the main products with impact in health. These compounds are energy source for colonic epithelium (butyrate) and peripheral tissues (acetate, propionate). The composition of microbiota and the diet carbohydrates determine the amount and proportion of SCFA produced²⁹. SCFA play a direct role on epithelial gut cells providing energy and promoting cell proliferation and differentiation. SCFA deficiency originates an energy deficit state that activates autophagy with impact in the integrity of intestinal barrier²⁷. SCFA receptors named G-protein coupled receptors (GPCR) are also present in peripheral tissues (white adipose tissue, skeletal muscle, and liver), acting as signaling molecules with impact in metabolism regulation (fatty acid oxidation, increasing leptin) and immune functions (reduction of pro-inflammatory cytokines and macrophage activation)^{30,31}.

Besides the production of SCFA, gut microbiota mediates other metabolism functions: bile acid conjugation to secondary bile acids acting as signaling molecules to regulate metabolism and immune cells³²; vitamin (complex B and K), amino acids and lipid synthesis, digestion or absorption^{27,33,34}; pathogens barrier²⁷; immune function by gut associated lymphoid tissue (GALT) that represents about 70% of the total immune system³⁵; neuroendocrine regulation by the gut-brain axis, where dysbalanced axis is associated to gastrointestinal diseases (chronic gut inflammations, pain, and metabolic disorders) as well as mood, behavior, stress disorders and satiety mechanism^{36,37}.

The intestinal microbiota is composed by numerous microorganisms mostly from Bacteria kingdom. About 90% belong to the phyla Firmicutes and Bacteroidetes and the most abundant genera are Bacteroides, Faecalibacterium and Bifidobacterium which proportions vary between individuals. At species level, there is a great variety which can originate a unique profile for each host³⁸.

Environmental variations influence intestinal microbiota composition such as pH, oxygen and nutrient availability. The bacterial concentration is higher in the lower portion of the gastrointestinal tract dominating the anaerobes (Figure 1).

Figure 1. Gut microbiota composition (Adapted from Gotteland³⁹ and Verdu et al.⁴⁰).



The composition of gut microbiota is defined at birth and evolves until about 3 years old, then it remains constant during lifetime and a few factors can temporarily change it such as antibiotics and diet^{39,41}. A human study found that in twins, the similarity of microbiota is within the same family members and not only in monozygotic twins. This fact indicates that environmental exposure has more impact than genotype in microbiota development⁴². Diet rich on non-digestible carbohydrates provide substrate to fermentation by gut bacteria. Limited amounts of fat and proteins are necessary to maintain a healthy gut metabolism. High fat and protein diets significantly reduced SCFA and alter intestinal bacteria composition^{43,44}. The supplementation with prebiotics promote the growth of specific gut microbiota species and the use of probiotics has benefits in host health, but the impact on large-term gut microbiota remains to be proven⁴⁵.

OBESITY AND GUT MICROBIOTA

Backhed et al.⁴⁶ inoculated germ-free mice with normal gut microbial cells from conventionally raised mice, which resulted in a 60% increase in body fat and insulin resistance, even with a 27% reduction in food intake. They also found that bacterial colonization increased the storage of triglycerides in the adipocytes of the inoculated mice⁴⁶. Ridaura et al.⁴⁷ also performed a study in which germ-free mice were colonized by fecal microbiota of twins discordant for obesity or by cultured collection of lean or obese animals. Fecal cultured or uncultured feces of obese animals originate significantly higher increase in body mass and adiposity than those of lean animals, whereas transplanted microbiota of lean animals was correlated to higher quantity of SCFA (butyrate and propionate). When the animals were cohoused (obese and lean) the obese animals stopped their body weight gain modifying also their microbiota profile like lean animals (increasing Bacteroides). The study was also conducted under 2 types of diet (low saturated fat and high saturated fat). The bacterial colonization and phenotype change only occur in low saturated fat diet revealing a diet-dependent mechanism between diet and microbiota⁴⁷.

Recently, increasingly studies embody the evidence of intestinal microbiota intervention in obesity. Intestinal dysbiosis implies an alteration in quality and quantity of intestinal commensal bacteria which means altered fermentation products (mainly SCFA), and occurs in obese people. In obesity, gut microbiota is known to be altered by a decreased ratio of Bacteroidetes to Firmicutes, with increased capacity to harvest energy from diet^{48,49}. It is also known that long-term diet habits influence composition of gut microbiota. Intestinal bacteria react to daily dietary fat and carbohydrates and change its metabolic pattern but the extent, mechanisms and consequences of a dietary shift are still unknown²⁹.

The main mechanisms influenced by SCFA regulation in peripheral tissues are well developed in several studies²⁹⁻³¹: energy harvesting^{50,51}, substrate metabolism⁵², energy expenditure^{53,54}, anorectic hormone production and appetite regulation⁵⁵⁻⁵⁸. These mechanisms may counterbalance the extra energy source that SCFA intestinal production represent in obesity as well as the establishment of the low-grade inflammatory state characteristic of obese persons²⁷. All these effects were observed in vitro or in animals, lacking evidence in humans, and so, those properties should be seen with caution.

FACTORS AFFECTING INTESTINAL COLONIZATION IN CHILDREN

The intestinal microbiota colonization occurs at birth, or before in uterus according to some authors, and it is the perinatal period the most important one to define gut microbiota in later ages.

Early life environment factors involved are⁵⁹: a) host genetics that controls gut microbiota diversity but, animal studies revealed that changes in diet population may alter gut microbiota despite host genetics^{60,61}; b) in uterus colonization were recent studies revealed a uterine microbiota in healthy pregnant women^{62,63}; c) maternal lifestyle including diet during pregnancy⁶⁴, overweight or excessive weight gain⁶⁵, and stress^{66,67} modulating gut microbiota, immune system and milk composition⁶⁸; d) birth delivery mode determine maternal transfer of vaginal, colonic and skin microbiota, colonizing the neonate specially with *Lactobacillus* and *Prevotella*⁶⁹⁻⁷¹; e) breastfeeding versus formula with a major impact on early microbiota composition and function, when compared to introduction of solid food or even the birth mode⁷⁰ and were milk bacteria (**specially bifidobacteria**) act as probiotics to children's gut⁷²⁻⁷⁵; f) solid food which increases diversity and promotes the growth of *Bacteroides* and *Clostridium* butyrate producers and may be a major determinant for gut microbiota development^{76,77}; g) antibiotics exposure that rapidly alters gut microbiota with short-term and long-term influences^{78,79}; h) hygiene level that also determines the microbial exposure and may influence the early development and diversity of gut microbiota⁸⁰⁻⁸²; i) prebiotics and probiotics administration to pregnant women and neonates have shown a modulation effect of child microbiota⁸³⁻⁸⁷. At 3 to 5 years old the gut microbiota composition is similar to adults and remains more or less stable. Changes may occur because of bacterial infections, surgeries, diet^{40,62}, lifestyle⁸⁸, and geographical area⁸⁹.

Continued research regarding the factors that can influence the development of human gut microbiota will enlighten the mechanism to achieve **and promote children's health**.

IMPACT OF GUT MICROBIOTA ON DEVELOPMENT OF CHILD OBESITY

Research in animal models have linked gut microbiota to obesity and is contributing to elucidate its mechanisms of interaction. However, there are not many studies in infants and several demand attention to their confounding factors undermining some results.

In Table 1 are summarized the studies in infant obesity related to gut microbiota.

Table 1. Studies on infant obesity and gut microbiome.

Type of study	N. of individuals	Ages (years)	Results (in obese individuals compared to lean)	Ref.
Case-study	20	4-5	- ↑ Enterobacteriaceae - ↓ Desulfovibrio and Akkermansia - ↔ Lactobacillus, Bifidobacterium and Bacteroides fragilis	90
Case-study	53	6-16	- ↑ Firmicutes: Bacteroidetes ratio - ↑ Lactobacillus spp.	91
Case-study	84	N.a.	- ↑ B. fragilis and Lactobacillus - ↓ Bifidobacterium	92
Case-study	15	8-14	- No differences in gut microbiota composition - ↑ SCFA butyrate and propionate	93
Prospective study	30	Followed until 10	- ↓ mother's colostrum adiponectin - ↓ Bifidobacterium at 3 months	94
Prospective study	138	Followed until 3	- ↑ B. fragilis and ↓ Staphylococcus between 3 weeks and 1 year	95
Prospective study	909	Followed until 3	- ↑ B. fragilis	96
Prospective study	246	Followed until 2	- ↓ Bacteroides spp. - ↑ Staphylococcus spp.	97
Follow-up interventional study (perinatal 1x10 ¹⁰ cfu of L. rhamnosus GG, ATCC 53103 against placebo)	113	Followed until 10	- Correlation found between perinatal gut microbiota modulation and early obesity until 48 months	98
randomized controlled trial (synbiotic against placebo for 8 weeks)	70	6-18	- significant decrease of tumor necrosis-α and interleukin-6, and significant increase in adiponectin; - No differences in C-reactive protein	99
Randomized controlled trial (prebiotic against placebo)	38	7-12	- normalized weight gain, reduced whole body and trunk body fat, modified primary fecal bile acids, and selectively altered gut microbiota	100
Open-labelled self-controlled nutritional intervention for 30 days	38	3-16	- both obese groups share the same dysbiosis - gut microbiota modulation within 30 days of non-digestible carbohydrates diet - significant decrease of inflammatory markers	101

↑: increase; ↓: decrease; ↔: maintenance; N.a.: non-available.

Observational studies reveal differences between gut microbiota of obese and lean children. Karlsson et al.⁹⁰ studied 20 overweight or obese children and 20 normal range children with ages between 4-5 years old. After analyzing their intestinal microbiota they found significant differences in abundance but only a tendency in their diversity. The abundance of Enterobacteriaceae was significantly higher in the obese or overweight children, whereas a significantly lower of *Desulfovibrio* and *Akkermansia muciniphila*-like bacteria. No significant differences were found in content of *Lactobacillus*, *Bifidobacterium* or the *B. fragilis* group⁹⁰. In another study comparing children between 6 and 16 years old the authors found elevated Firmicutes-to-Bacteroidetes ratio in obese children compared with lean ones. Additionally, low relative proportions of *B. vulgatus* and high levels of *Lactobacillus* spp. were observed in the obese children⁹¹. Gut microbiota of 30 obese, 24 overweight and 30 lean children were verified, and the authors found a positive correlation between BMI and high levels of *B. fragilis* group and *Lactobacillus* spp. while a negative correlation was found for *Bifidobacterium* spp.⁹². Obese (n=15) and normal weight (n=15) children aged between 8 and 14 years old were studied for their gut microbiota showing no significant quantitative differences in gut microbiota. However, higher concentrations of butyrate and propionate were found in obese versus normal weight children. Lower concentrations of intermediate metabolites detected in obese children, may suggest higher metabolic activity by obese gut microbiota leading to future dysbiosis⁹³.

In a prospective study, Luoto et al.⁹⁴ correlated the post-natal diet (maternal colostrum adiponectin concentration) and gut microbiota (at the age of 3 months) to subsequently normal weight (n=15) versus overweight (n=15) 10 years old children. Sex, gestational age, BMI at birth, mode of delivery, probiotic intervention, and duration of breast-feeding were similar in both groups. Colostrum adiponectin concentrations were significantly higher in mothers whose children were normal weight as well as the *Bifidobacterium* levels⁹⁴. In a similar study design, 138 infants were studied for their gut microbiota at 3, 26 and 52 weeks of age and related to their BMI at 1 and 3 years old. A low *Staphylococcus* and a high *B. fragilis* concentration, was associated with a higher BMI during the first three years of life⁹⁵. The same correlation was found between *B. fragilis* group and a higher BMI when fecal samples of 909 one-month-old infants were analyzed and BMI were evaluated between 1 and 10 years old⁹⁶.

An interesting prospective study tried to establish a timeline between early gut microbiota patterns and infant growth. Collection of fecal samples were made at postpartum day 4 (mother sample) and their infants at 4, 10, 30, and 120 days old, totalizing 246 children after the study of inclusion process was concluded. Possible study confounders such as antibiotics use (after day 4 of life), sex, having received milk substitutes, maternal smoking, and parity were analysed and

removed. The aim of this work was the detection of specific gut microbiota groups that were significantly associated with infant growth trajectory. The samples showed 16 gut ecosystem developing patterns that were detected over time and some results were: detection of *Bacteroides* spp. at day 30 was significantly associated with reducing growth in males when compared to non-detection; detection of *Staphylococcus* spp. at day 4 was associated with expected growth in females and males; *Escherichia coli* detection from day 4 through to 30 was associated with expected growth in males. These results may be an insight to establish a correlation between changes in gut microbiota development and consequent risk of obesity. This work also developed a novel approach to provide a potential time-dependent exposure window by observational data otherwise only occurred by experimental data⁹⁷.

Nadal et al.¹⁰² and Santacruz et al.¹⁰³ found gut microbiota changes in obese adolescents when they altered their lifestyle, mainly diet and exercise, suggesting interactions between diet, gut microbiota and host metabolism and immunity in obesity.

To better establish a timeline between early gut microbiota and its impact in developing obesity later in life, more experimental studies need to be done. Very few can be found since ethical issues limit their elaboration. Luoto et al.⁹⁸ performed an interventional study where 159 pregnant women were randomized and double-blinded to receive probiotics (1×10^{10} CFU of *L. rhamnosus* GG, ATCC 53103) or placebo 4 weeks before delivery and extended until 6 months after. 113 children were enrolled in the study and their anthropometric measures were taken at 3, 6, 12 and 24 months and at 4, 7 and 10 years. The results showed that the perinatal probiotic intervention appeared to moderate the initial phase of excessive weight gain (until 24-48 months), especially among children who later became overweight, but not the second phase of excessive weight gain (after 4 years old). Early gut microbiota modulation appears to influence only infant growth in the first years of life⁹⁸.

A symbiotic (Protexin® 2.0×10^8 CFU/day of *L. casei*, *L. rhamnosus*, *Streptococcus thermophilus*, *B. breve*, *L. acidophilus*, *B. longum*, *L. delbrueckii* subsp. *bulgaricus*, and fructo-oligosaccharides, vitamin E, vitamin A, and vitamin C) was tested against placebo in a group of obese children of 6-18 years old in a 8 week randomized controlled trial in order to study its impact in obesity inflammatory markers. The symbiotic group had significant decrease in values of tumor necrosis- α and interleukin-6, with significant increase in adiponectin. No differences were found in C-reactive protein. The results were depending on weight reduction. Considering the duration of the intervention, symbiotic supplementation may positively influence inflammation markers⁹⁹.

In a randomized controlled trial¹⁸, obese children aged between 7 and 12 years were randomly included to receive a prebiotic oligofructose-enriched inulin (n=20) or a placebo for 16 weeks. No

significant differences were found in BMI of prebiotic group while BMI significantly increased in the placebo group. Percent of total body fat was significantly lowered with prebiotic compared with placebo showing differences in body distribution. Lean mass had a significant increase in both groups. It was also observed a decrease tendency in inflammatory markers in prebiotic group. In respect to gut microbiota modulation it was observed an increase of *Bifidobacterium* spp. in the prebiotic group. The prebiotic administration normalized weight gain, reduced whole body and trunk body fat, modified primary fecal bile acids (microbiota metabolites), and selectively altered gut microbiota¹⁰⁰.

Zhang et al.¹⁰¹ performed a nutritional intervention in 38 hospitalized children (3-16 years old) suffering from genetic obesity (n=17) or common obesity (n=21). After a diet rich in fermentable non-digestible carbohydrates (whole grains, vegetable, fruits, nuts, traditional Chinese medicinal food plants and prebiotics) for 30 days a significant weight loss and changes of the gut microbiota were observed together with a reduction of metabolic deterioration and inflammation markers (C-reactive protein, serum amyloid A protein, α -acid glycoprotein and white blood cell count). The levels of adiponectin increased and leptin decreased. Lipopolysaccharide binding protein, a marker for bacterial antigen load in the blood also decreased. The study revealed that both groups of obese children shared the same pattern of dysbiosis and in both it was observed gut microbiota modulation after the intervention. The authors also performed an in vivo essay by faecal transplantation to germ-free wild-type C57BL/6J mice. Those who received pre-intervention samples developed higher fat mass and presented high inflammatory markers and those who received post-intervention samples remained with the normal weight. The significant change on clinical parameters suggests overall structural changes at individual microbiome level. They also proved that long-term gut microbiota modulation by diet can be done¹⁰¹.

DISCUSSION

Results from literature are often inconsistent as many confounders exist, namely the fact that different microbe identification techniques are used and the intra-individual and genotype differences that naturally occur. Besides in vivo animal essays, human clinical trial results are controversial and scarce in infant population.

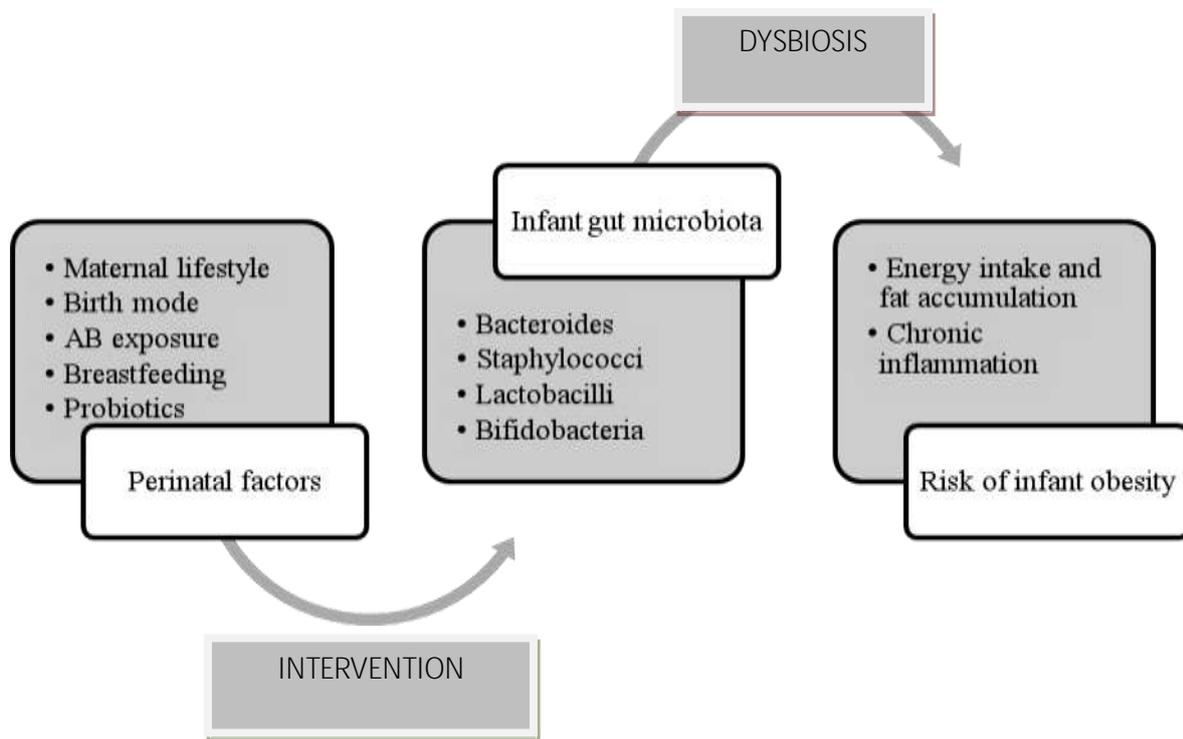
Until now it is not clear enough the correlation between gut microbiota and obesity but some statements can be summarized: gut microbiota is developed in early life and several factors contribute to its composition; gut microbiota from obese individuals differ from healthy ones, it is also apparent that *Bacteroides* spp. seem to play an important role in regulating childhood microbiota as well as lactobacilli, bifidobacteria, staphylococci, and a low *Bacteroides*/*Firmicutes* ratio, which can influence the development of overweight later in life.

Scientists are increasingly agreeing that dysbiosis can be one of the causes of obesity and that diet and prebiotic/probiotic interventions can qualitatively change gut microbiota during a period of time that is yet unknown. It is also well accepted that shifts in microbiota may result in healthier clinical parameters for obese individuals. Nevertheless, despite the intense research seen in the recent years, there is still much work to do either in animal models to elucidate the molecular mechanisms in which the gut microbiota controls weight gain, in identifying specific microbe species for shaping body composition or in laboratory analysis to standardize microbe identification and even in well-design human (infant) studies aiming to clarify the 'healthy gut microbiota' and which measures are effective to modulate a healthy gut microbiota development. Several recent reviews conclude the positive role of gut microbiota modulation in adult obesity treatment¹⁰⁴⁻¹⁰⁶. However, there are no sufficient data to state that child dysbiosis increase the risk of obesity later in life besides a few studies reported some discordant changes in gut microbiota of obese children. A timeline establishment between early gut microbiota and obesity was attempted by Richard A. et al.⁹⁷ but the study was very limited in time.

Even if more studies are still needing to clearly claim that gut microbiota modulation will play an important role in the treatment of obesity, earlier interventional trials in obese children open promising doors in that direction.

It is already known that pregnancy and perinatal factors (maternal health and lifestyle, birth mode, breastfeeding) are fundamental to the development of infant gut microbiota. Infant dysbiosis may be transmitted and responsible for obesity programming. Primary prevention strategies may include modeling maternal and infants gut microbiota and break the obesity cycle. The use of prebiotics, probiotics and changes on maternal lifestyle may be successful interventions to avoid children dysbiosis and consequent obesity (Figure 2).

Figure 2. Gut microbiota development and risk of infant obesity.



CONCLUSIONS

Despite the large amount of scientific publications, there is still much work to do regarding the clarification of mechanisms and the possible therapy for childhood obesity.

COMPETING INTERESTS

Authors state that there are no conflicts of interest in preparing the manuscript.

REFERENCES

- (1) World Health Organization. Obesity and overweight. Fact Sheet [Internet]. World Health Organization. 2016 [citado 1 de septiembre de 2016]. Disponible en: <http://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight>
- (2) Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2014;384(9945):766-81.
- (3) Herrera BM, Keildson S, Lindgren CM. Genetics and epigenetics of obesity. *Maturitas*. 2011;69(1):41-9.
- (4) Katzmarzyk PT, Barlow S, Bouchard C, Catalano PM, Hsia DS, Inge TH, et al. An evolving scientific basis for the prevention and treatment of pediatric obesity. *Int J Obes*. 2014;38(7):887-905.
- (5) Woo Baidal JA, Locks LM, Cheng ER, Blake-Lamb TL, Perkins ME, Taveras EM. Risk Factors for Childhood Obesity in the First 1,000 Days: A Systematic Review. *Am J Prev Med*. 2016;50(6):761-79.
- (6) Albuquerque D, Stice E, Rodríguez-López R, Manco L, Nóbrega C. Current review of genetics of human obesity: from molecular mechanisms to an evolutionary perspective. *Mol Genet Genomics*. 2015;290(4):1191-221.
- (7) Goldstone AP, Beales PL. Genetic obesity syndromes. *Front Horm Res*. 2008;36:37-60.
- (8) Chesi A, Grant SFA. The Genetics of Pediatric Obesity. *Trends Endocrinol Metab*. 2015;26(12):711-21.
- (9) Hinney A, Vogel CIG, Hebebrand J. From monogenic to polygenic obesity: recent advances. *Eur Child Adolesc Psychiatry*. 2010;19(3):297-310.
- (10) Young BE, Johnson SL, Krebs NF. Biological determinants linking infant weight gain and child obesity: current knowledge and future directions. *Adv Nutr*. 2012;3(5):675-86.
- (11) Gillman MW. The first months of life: a critical period for development of obesity. *Am J Clin Nutr*. 2008;87(6):1587-9.
- (12) Thompson AL. Developmental origins of obesity: early feeding environments, infant growth, and the intestinal microbiome. *Am J Hum Biol*. 2012;24(3):350-60.
- (13) Kon IY, Shilina NM, Gmoshinskaya MV, Ivanushkina TA. The study of breast milk IGF-1, leptin, ghrelin and adiponectin levels as possible reasons of high weight gain in breast-fed infants. *Ann Nutr Metab*. 2014;65(4):317-23.

- (14) Brunner S, Schmid D, Zang K, Much D, Knoeferl B, Kratzsch J, et al. Breast milk leptin and adiponectin in relation to infant body composition up to 2 years. *Pediatr Obes.* 2015;10(1):67-73.
- (15) Dewey KG. Growth characteristics of breast-fed compared to formula-fed infants. *Biol Neonate.* 1998;74(2):94-105.
- (16) Weng SF, Redsell SA, Swift JA, Yang M, Glazebrook CP. Systematic review and meta-analyses of risk factors for childhood overweight identifiable during infancy. *Arch Dis Child.* 2012;97(12):1019-26.
- (17) Grunewald M, Hellmuth C, Demmelmair H, Koletzko B. Excessive weight gain during full breast-feeding. *Ann Nutr Metab.* 2014;64(3-4):271-5.
- (18) Fenton TR, Premji SS, Al-Wassia H, Sauve RS. Higher versus lower protein intake in formula-fed low birth weight infants. *Cochrane Database Syst Rev.* 2014;(4):CD003959.
- (19) Castillo-Laura H, Santos IS, Quadros LCM, Matijasevich A. Maternal obesity and offspring body composition by indirect methods: a systematic review and meta-analysis. *Cad Saude Publica.* 2015;31(10):2073-92.
- (20) Ahuja S, Boylan M, Hart SL, Román-Shriver C, Spallholz JE, Pence BC, et al. **Glucose and Insulin Levels are Increased in Obese and Overweight Mothers' Breast-Milk.** *Food Nutr Sci.* 2011;2(3):201-6.
- (21) Fleddermann M, Demmelmair H, Grote V, Nikolic T, Trisic B, Koletzko B. Infant formula composition affects energetic efficiency for growth: the BeMIM study, a randomized controlled trial. *Clin Nutr.* 2014;33(4):588-95.
- (22) Weber M, Grote V, Closa-Monasterolo R, Escribano J, Langhendries J-P, Dain E, et al. Lower protein content in infant formula reduces BMI and obesity risk at school age: follow-up of a randomized trial. *Am J Clin Nutr.* 2014;99(5):1041-51.
- (23) Inostroza J, Haschke F, Steenhout P, Grathwohl D, Nelson SE, Ziegler EE. Low-protein formula slows weight gain in infants of overweight mothers. *J Pediatr Gastroenterol Nutr.* 2014;59(1):70-7.
- (24) Oddy WH. Infant feeding and obesity risk in the child. *Breastfeed Rev.* 2012;20(2):7-12.
- (25) Klag EA, McNamara K, Geraghty SR, Keim SA. Associations Between Breast Milk Feeding, Introduction of Solid Foods, and Weight Gain in the First 12 Months of Life. *Clin Pediatr.* 2015;54(11):1059-67.
- (26) Sabin MA, Kiess W. Childhood obesity: Current and novel approaches. *Best Pract Res Clin Endocrinol Metab.* 2015;29(3):327-38.

- (27) Brestoff JR, Artis D. Commensal bacteria at the interface of host metabolism and the immune system. *Nat Immunol.* 2013;14(7):676-84.
- (28) Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell.* 2012;148(6):1258-70.
- (29) Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature.* 2012;489(7415):242-9.
- (30) Kasubuchi M, Hasegawa S, Hiramatsu T, Ichimura A, Kimura I. Dietary gut microbial metabolites, short-chain fatty acids, and host metabolic regulation. *Nutrients.* 2015;7(4):2839-49.
- (31) Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol.* 2015;11(10):577-91.
- (32) Duboc H, Rajca S, Rainteau D, Benarous D, Maubert M-A, Quervain E, et al. Connecting dysbiosis, bile-acid dysmetabolism and gut inflammation in inflammatory bowel diseases. *Gut.* 2013;62(4):531-9.
- (33) Claus SP, Ellero SL, Berger B, Krause L, Bruttin A, Molina J, et al. Colonization-induced host-gut microbial metabolic interaction. *MBio.* 2011;2(2):e00271-00210.
- (34) Matsumoto M, Kibe R, Ooga T, Aiba Y, Kurihara S, Sawaki E, et al. Impact of intestinal microbiota on intestinal luminal metabolome. *Sci Rep.* 2012;2:233.
- (35) Vighi G, Marcucci F, Sensi L, Di Cara G, Frati F. Allergy and the gastrointestinal system. *Clin Exp Immunol.* 2008;153(Suppl 1):3-6.
- (36) Moloney RD, Desbonnet L, Clarke G, Dinan TG, Cryan JF. The microbiome: stress, health and disease. *Mamm Genome.* 2014;25(1-2):49-74.
- (37) Morton GJ, Meek TH, Schwartz MW. Neurobiology of food intake in health and disease. *Nat Rev Neurosci.* 2014;15(6):367-78.
- (38) Robles-Alonso V, Guarner F. Progreso en el conocimiento de la microbiota intestinal humana. *Nutr Hosp.* 2013;28(3):553-7.
- (39) Gotteland M. El papel de la microbiota intestinal en el desarrollo de la obesidad y de la diabetes de tipo-2. *Rev Chil Endocrinol Diabetes.* 2013;6(4):155-62.
- (40) Verdu EF, Galipeau HJ, Jabri B. Novel players in coeliac disease pathogenesis: role of the gut microbiota. *Nat Rev Gastroenterol Hepatol.* 2015;12(9):497-506.
- (41) Soderborg TK, Borengasser SJ, Barbour LA, Friedman JE. Microbial transmission from mothers with obesity or diabetes to infants: an innovative opportunity to interrupt a vicious cycle. *Diabetologia.* 2016;59(5):895-906.

- (42) Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JI. The human microbiome project. *Nature*. 2007;449(7164):804-10.
- (43) Brinkworth GD, Noakes M, Clifton PM, Bird AR. Comparative effects of very low-carbohydrate, high-fat and high-carbohydrate, low-fat weight-loss diets on bowel habit and faecal short-chain fatty acids and bacterial populations. *Br J Nutr*. 2009;101(10):1493-502.
- (44) Russell WR, Gratz SW, Duncan SH, Holtrop G, Ince J, Scobbie L, et al. High-protein, reduced-carbohydrate weight-loss diets promote metabolite profiles likely to be detrimental to colonic health. *Am J Clin Nutr*. 2011;93(5):1062-72.
- (45) Scott KP, Gratz SW, Sheridan PO, Flint HJ, Duncan SH. The influence of diet on the gut microbiota. *Pharmacol Res*. 2013;69(1):52-60.
- (46) Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA*. 2004;101(44):15718-23.
- (47) Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science*. 2013;341(6150):1241214.
- (48) Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*. 2006;444(7122):1027-31.
- (49) López-Cepero AA, Palacios C. Association of the Intestinal Microbiota and Obesity. *P R Health Sci J*. 2015;34(2):60-4.
- (50) Fernandes J, Su W, Rahat-Rozenbloom S, Wolever TMS, Comelli EM. Adiposity, gut microbiota and faecal short chain fatty acids are linked in adult humans. *Nutr Diabetes*. 2014;4:e121.
- (51) Turnbaugh PJ, Hamady M, Yatsunencko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature*. 2009;457(7228):480-4.
- (52) De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchamp A, et al. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell*. 2014;156(1-2):84-96.
- (53) Donohoe DR, Garge N, Zhang X, Sun W, O'Connell TM, Bunger MK, et al. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab*. 2011;13(5):517-26.
- (54) Velagapudi VR, Hezaveh R, Reigstad CS, Gopalacharyulu P, Yetukuri L, Islam S, et al. The gut microbiota modulates host energy and lipid metabolism in mice. *J Lipid Res*. 2010;51(5):1101-12.

- (55) Frost G, Sleeth ML, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, et al. The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. *Nat Commun.* 2014;5:3611.
- (56) Zhou J, Martin RJ, Tulley RT, Raggio AM, McCutcheon KL, Shen L, et al. Dietary resistant starch upregulates total GLP-1 and PYY in a sustained day-long manner through fermentation in rodents. *Am J Physiol Endocrinol Metab.* 2008;295(5):E1160-1166.
- (57) Cani PD, Lecourt E, Dewulf EM, Sohet FM, Pachikian BD, Naslain D, et al. Gut microbiota fermentation of prebiotics increases satietogenic and incretin gut peptide production with consequences for appetite sensation and glucose response after a meal. *Am J Clin Nutr.* 2009;90(5):1236-43.
- (58) Myers MG, Leibel RL, Seeley RJ, Schwartz MW. Obesity and leptin resistance: distinguishing cause from effect. *Trends Endocrinol Metab.* 2010;21(11):643-51.
- (59) Chan YK, Estaki M, Gibson DL. Clinical consequences of diet-induced dysbiosis. *Ann Nutr Metab.* 2013;63(Suppl 2):28-40.
- (60) Spor A, Koren O, Ley R. Unravelling the effects of the environment and host genotype on the gut microbiome. *Nat Rev Microbiol.* 2011;9(4):279-90.
- (61) Zhang C, Zhang M, Wang S, Han R, Cao Y, Hua W, et al. Interactions between gut microbiota, host genetics and diet relevant to development of metabolic syndromes in mice. *ISME J.* 2010;4(2):232-41.
- (62) Rodríguez JM, Murphy K, Stanton C, Ross RP, Kober OI, Juge N, et al. The composition of the gut microbiota throughout life, with an emphasis on early life. *Microb Ecol Health Dis.* 2015;26:26050.
- (63) Aagaard K, Ma J, Antony KM, Ganu R, Petrosino J, Versalovic J. The placenta harbors a unique microbiome. *Sci Transl Med.* 2014;6(237):237ra65.
- (64) Sonnenburg ED, Smits SA, Tikhonov M, Higginbottom SK, Wingreen NS, Sonnenburg JL. Diet-induced extinctions in the gut microbiota compound over generations. *Nature.* 2016;529(7585):212-5.
- (65) Collado MC, Isolauri E, Laitinen K, Salminen S. Distinct composition of gut microbiota during pregnancy in overweight and normal-weight women. *Am J Clin Nutr.* 2008;88(4):894-9.
- (66) Jašarević E, Rodgers AB, Bale TL. A novel role for maternal stress and microbial transmission in early life programming and neurodevelopment. *Neurobiol Stress.* 2015;1:81-8.

- (67) Zijlmans MAC, Korpela K, Riksen-Walraven JM, de Vos WM, de Weerth C. Maternal prenatal stress is associated with the infant intestinal microbiota. *Psychoneuroendocrinology*. 2015;53:233-45.
- (68) Collado MC, Laitinen K, Salminen S, Isolauri E. Maternal weight and excessive weight gain during pregnancy modify the immunomodulatory potential of breast milk. *Pediatr Res*. 2012;72(1):77-85.
- (69) Jakobsson HE, Abrahamsson TR, Jenmalm MC, Harris K, Quince C, Jernberg C, et al. Decreased gut microbiota diversity, delayed Bacteroidetes colonisation and reduced Th1 responses in infants delivered by caesarean section. *Gut*. 2014;63(4):559-66.
- (70) Bäckhed F, Roswall J, Peng Y, Feng Q, Jia H, Kovatcheva-Datchary P, et al. Dynamics and Stabilization of the Human Gut Microbiome during the First Year of Life. *Cell Host Microbe*. 2015;17(5):690-703.
- (71) Dominguez-Bello MG, De Jesus-Laboy KM, Shen N, Cox LM, Amir A, Gonzalez A, et al. Partial restoration of the microbiota of cesarean-born infants via vaginal microbial transfer. *Nat Med*. 2016;22(3):250-3.
- (72) Fernández L, Langa S, Martín V, Maldonado A, Jiménez E, Martín R, et al. The human milk microbiota: origin and potential roles in health and disease. *Pharmacol Res*. 2013;69(1):1-10.
- (73) Favier CF, Vaughan EE, De Vos WM, Akkermans ADL. Molecular monitoring of succession of bacterial communities in human neonates. *Appl Environ Microbiol*. 2002;68(1):219-26.
- (74) Perez PF, Doré J, Leclerc M, Levenez F, Benyacoub J, Serrant P, et al. Bacterial imprinting of the neonatal immune system: lessons from maternal cells? *Pediatrics*. 2007;119(3):e724-732.
- (75) Bode L. Human milk oligosaccharides: every baby needs a sugar mama. *Glycobiology*. 2012;22(9):1147-62.
- (76) Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, et al. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci USA*. 2011;108(Suppl 1):4578-85.
- (77) Laursen MF, Andersen LBB, Michaelsen KF, Mølgaard C, Trolle E, Bahl MI, et al. Infant Gut Microbiota Development Is Driven by Transition to Family Foods Independent of Maternal Obesity. *mSphere*. 2016;1(1):00069-15.
- (78) Francino MP. Antibiotics and the Human Gut Microbiome: Dysbioses and Accumulation of Resistances. *Front Microbiol*. 2015;6:1543.
- (79) Kumar H, Rautava S, Collado M, Borzykh N, Loyttyniemi E, Isolauri E, et al. Neonatal Antibiotic Exposure Alters Compositional Gut Microbiota Development During the First 6 Months of Life. *FASEB J*. 2015;29(Suppl. 1):1.

- (80) Ege MJ, Mayer M, Normand A-C, Genuneit J, Cookson WOCM, Braun-Fahrlander C, et al. Exposure to environmental microorganisms and childhood asthma. *N Engl J Med.* 2011;364(8):701-9.
- (81) Zhou D. Impact of sanitary living environment on gut microbiota. *Precis Med.* 2016;2:e1161.
- (82) Azad MB, Konya T, Maughan H, Guttman DS, Field CJ, Sears MR, et al. Infant gut microbiota and the hygiene hypothesis of allergic disease: impact of household pets and siblings on microbiota composition and diversity. *Allergy Asthma Clin Immunol.* 2013;9(1):15.
- (83) Sanchez M, Panahi S, Tremblay A. Childhood obesity: a role for gut microbiota? *Int J Environ Res Public Health.* 2015;12(1):162-75.
- (84) Rautava S, Collado MC, Salminen S, Isolauri E. Probiotics modulate host-microbe interaction in the placenta and fetal gut: a randomized, double-blind, placebo-controlled trial. *Neonatology.* 2012;102(3):178-84.
- (85) Luoto R, Laitinen K, Nermes M, Isolauri E. Impact of maternal probiotic-supplemented dietary counselling on pregnancy outcome and prenatal and postnatal growth: a double-blind, placebo-controlled study. *Br J Nutr.* 2010;103(12):1792-9.
- (86) Gueimonde M, Sakata S, Kalliomäki M, Isolauri E, Benno Y, Salminen S. Effect of maternal consumption of lactobacillus GG on transfer and establishment of fecal bifidobacterial microbiota in neonates. *J Pediatr Gastroenterol Nutr.* 2006;42(2):166-70.
- (87) Abdulkadir B, Nelson A, Skeath T, Marrs ECL, Perry JD, Cummings SP, et al. Routine Use of Probiotics in Preterm Infants: Longitudinal Impact on the Microbiome and Metabolome. *Neonatology.* 2016;109(4):239-47.
- (88) Mika A, Fleshner M. Early-life exercise may promote lasting brain and metabolic health through gut bacterial metabolites. *Immunol Cell Biol.* 2016;94(2):151-7.
- (89) Suzuki TA, Worobey M. Geographical variation of human gut microbial composition. *Biol Lett.* 2014;10(2):20131037.
- (90) Karlsson CLJ, Onnerfält J, Xu J, Molin G, Ahrné S, Thorngren-Jerneck K. The microbiota of the gut in preschool children with normal and excessive body weight. *Obesity.* 2012;20(11):2257-61.
- (91) Bervoets L, Van Hoorenbeeck K, Kortleven I, Van Noten C, Hens N, Vael C, et al. Differences in gut microbiota composition between obese and lean children: a cross-sectional study. *Gut Pathog.* 2013;5(1):10.

- (92) Ignacio A, Fernandes MR, Rodrigues V a. A, Groppo FC, Cardoso AL, Avila-Campos MJ, et al. Correlation between body mass index and faecal microbiota from children. *Clin Microbiol Infect.* 2016;22(3):258.e1-8.
- (93) Payne AN, Chassard C, Zimmermann M, Müller P, Stinca S, Lacroix C. The metabolic activity of gut microbiota in obese children is increased compared with normal-weight children and exhibits more exhaustive substrate utilization. *Nutr Diabetes.* 2011;1:e12.
- (94) Luoto R, Kalliomäki M, Laitinen K, Delzenne NM, Cani PD, Salminen S, et al. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J Pediatr Gastroenterol Nutr.* 2011;52(1):90-5.
- (95) Vael C, Verhulst SL, Nelen V, Goossens H, Desager KN. Intestinal microflora and body mass index during the first three years of life: an observational study. *Gut Pathog.* 2011;3(1):8.
- (96) Scheepers LEJM, Penders J, Mbakwa CA, Thijs C, Mommers M, Arts ICW. The intestinal microbiota composition and weight development in children: the KOALA Birth Cohort Study. *Int J Obes.* 2015;39(1):16-25.
- (97) White RA, Bjørnholt JV, Baird DD, Midtvedt T, Harris JR, Pagano M, et al. Novel developmental analyses identify longitudinal patterns of early gut microbiota that affect infant growth. *PLoS Comput Biol.* 2013;9(5):e1003042.
- (98) Luoto R, Kalliomäki M, Laitinen K, Isolauri E. The impact of perinatal probiotic intervention on the development of overweight and obesity: follow-up study from birth to 10 years. *Int J Obes.* 2010;34(10):1531-7.
- (99) Kelishadi R, Farajian S, Safavi M, Mirlohi M, Hashemipour M. A randomized triple-masked controlled trial on the effects of synbiotics on inflammation markers in overweight children. *J Pediatr.* 2014;90(2):161-8.
- (100) Nicolucci AC, Hume MP, Martínez I, Mayengbam S, Walter J, Reimer RA. Prebiotics Reduce Body Fat and Alter Intestinal Microbiota in Children Who Are Overweight or With Obesity. *Gastroenterology.* 2017;153(3):711-22.
- (101) Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, et al. Dietary Modulation of Gut Microbiota Contributes to Alleviation of Both Genetic and Simple Obesity in Children. *EBioMedicine.* 2015;2(8):968-84.
- (102) Nadal I, Santacruz A, Marcos A, Warnberg J, Garagorri JM, Garagorri M, et al. Shifts in clostridia, bacteroides and immunoglobulin-coating fecal bacteria associated with weight loss in obese adolescents. *Int J Obes.* 2009;33(7):758-67.

- (103) Santacruz A, Marcos A, Wärnberg J, Martí A, Martín-Matillas M, Campoy C, et al. Interplay between weight loss and gut microbiota composition in overweight adolescents. *Obesity*. 2009;17(10):1906-15.
- (104) Seganfredo FB, Blume CA, Moehlecke M, Giongo A, Casagrande DS, Spolidoro JVN, et al. Weight-loss interventions and gut microbiota changes in overweight and obese patients: a systematic review. *Obes Rev*. 2017;18(8):832-51.
- (105) Dahiya DK, Renuka null, Puniya M, Shandilya UK, Dhewa T, Kumar N, et al. Gut Microbiota Modulation and Its Relationship with Obesity Using Prebiotic Fibers and Probiotics: A Review. *Front Microbiol*. 2017;8:563.
- (106) Li J, Riaz Rajoka MS, Shao D, Jiang C, Jin M, Huang Q, et al. Strategies to increase the efficacy of using gut microbiota for the modulation of obesity. *Obes Rev*. 2017;18(11):1260-71.